



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵ :

A61L 29/00, 27/00, 31/00

A61L 15/44, A61M 35/00

A1

(11) International Publication Number:

WO 92/11043

(43) International Publication Date:

9 July 1992 (09.07.92)

(21) International Application Number: PCT/CA91/00453

(22) International Filing Date: 23 December 1991 (23.12.91)

(30) Priority data:

2,033,107

24 December 1990 (24.12.90) CA

(71) Applicant (for all designated States except US): Q-LIFE SYSTEMS, INC. [CA/CA]; 1180 Clyde Court, Kingston, Ontario K7P 2E4 (CA).

(72) Inventors; and

(75) Inventors/Applicants (for US only): BURRELL, Robert, Edward [CA/CA]; 651 Braeside Crescent, Kingston, Ontario K7P 1G6 (CA). ROSENFELD, Aron, Marcus [CA/CA]; 708-64 Ontario Street, Kingston, Ontario K7L 5J4 (CA). SMITH, Timothy, J., N. [CA/CA]; 21 Pickwick Place, Kingston, Ontario K7M 1M1 (CA).

(74) Agent: SCOTT & AYLEN; 60 Queen Street, Ottawa, Ontario K1P 5Y7 (CA).

(81) Designated States: AT (European patent), AU, BE (European patent), CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, LU (European patent), MC (European patent), NL (European patent), SE (European patent), US.

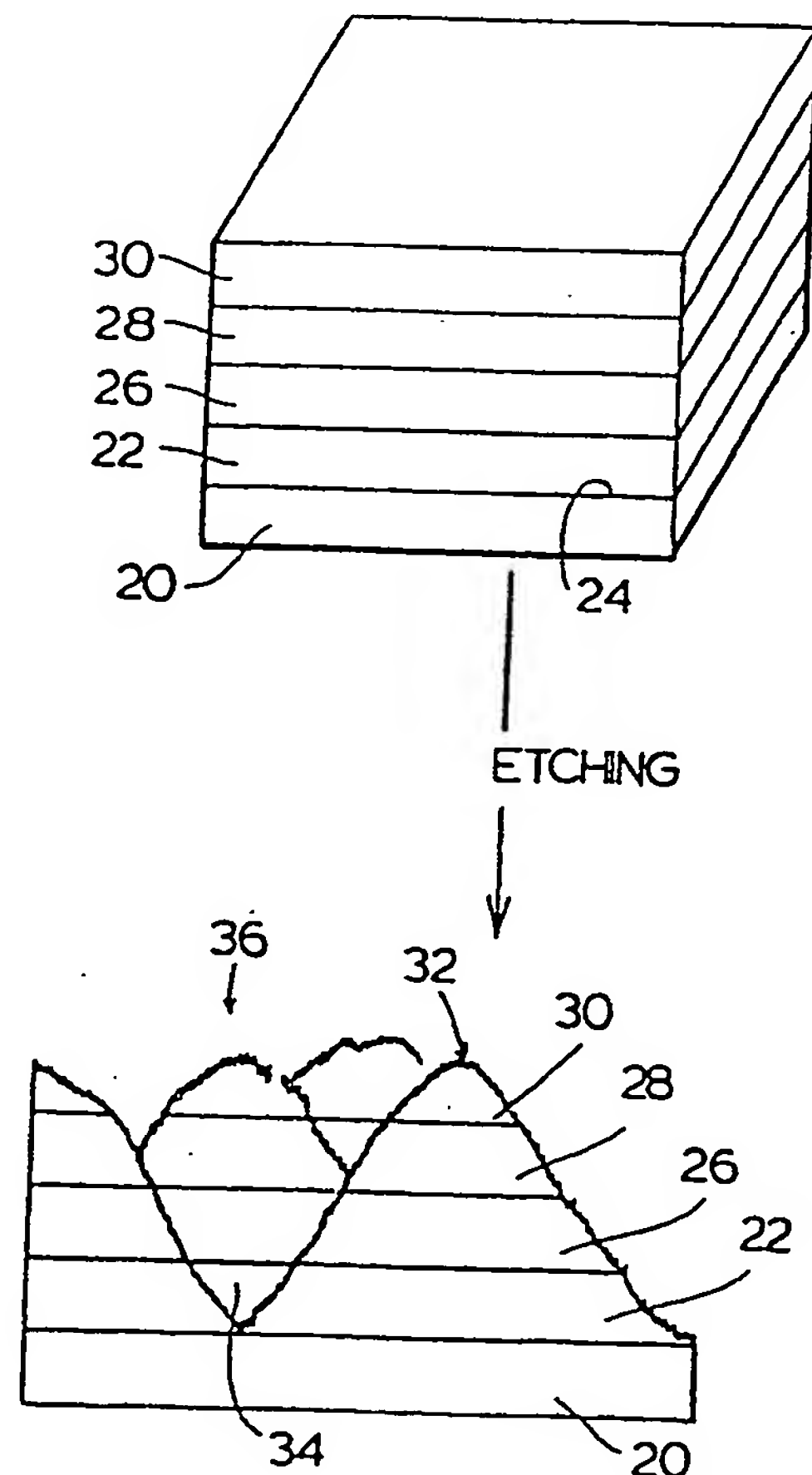
Published

With international search report.

(54) Title: ACTIVELY STERILE SURFACES

(57) Abstract

An actively antimicrobial surface for a substrate and for use in a biologically dynamic environment, such as for treating and preventing microbial infections, comprises a film consisting of at least an antimicrobial element (22, 26) and another electrochemically nobler element (26, 22) and which forms multitudinous galvanic cells with electrolyte containing biological fluids, such as body fluids from wounds, etc., for releasing the antimicrobial element at the surface.



FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	ES	Spain	MG	Madagascar
AU	Australia	FI	Finland	ML	Mali
BB	Barbados	FR	France	MN	Mongolia
BE	Belgium	GA	Gabon	MR	Mauritania
BF	Burkina Faso	GB	United Kingdom	MW	Malawi
BG	Bulgaria	GN	Guinea	NL	Netherlands
BJ	Benin	GR	Greece	NO	Norway
BR	Brazil	HU	Hungary	PL	Poland
CA	Canada	IT	Italy	RO	Romania
CF	Central African Republic	JP	Japan	SD	Sudan
CG	Congo	KP	Democratic People's Republic of Korea	SE	Sweden
CH	Switzerland	KR	Republic of Korea	SN	Senegal
CI	Côte d'Ivoire	LI	Liechtenstein	SU ⁺	Soviet Union
CM	Cameroon	LK	Sri Lanka	TD	Chad
CS	Czechoslovakia	LU	Luxembourg	TG	Togo
DE*	Germany	MC	Monaco	US	United States of America
DK	Denmark				

⁺ Any designation of "SU" has effect in the Russian Federation. It is not yet known whether any such designation has effect in other States of the former Soviet Union.

- 1 -

Actively Sterile SurfacesTechnical Field

This invention relates to actively antimicrobial surfaces
5 useful in avoiding, preventing and treating bacterial, fungal and
microbial infections generally by releasing substances which are
active in suppressing such organisms.

It has been appreciated for some time that certain ions and
compounds are very effective antimicrobials for treating and/or
10 killing bacterial and fungal entities. Metallic silver and silver
salts have been used to inhibit growth of microorganisms on fresh
wounds and the like. Silver nitrates have been commonly used as
bactericidal agents.

A problem with the in vivo use of metal for metal ion
15 therapy in preventing and treating infections is that the results
have never been too spectacular. This is primarily due to the very
low concentration of active metal ions and, in most situations, the
rapid decline in the presence of metal ions when derived from salt
solutions as administered to the infected area.

20

Background Art

Considerable research work has been conducted in the field
of silver metal treatment of bacterial infections. A variety of
silver coated nylon cloths and fibers have been investigated, such as
25 disclosed in Antimicrobial Agents and Chemotherapy, March 1983,
pp. 356-359, Deitch et al, "Silver-Nylon: a New Antimicrobial
Agent"; Antimicrobial Agents and Chemotherapy, January 1987, pp.
93-99, MacKeen et al, "Silver-Coated Nylon Fiber as an Antibacterial
Agent"; and in The Journal of Trauma, Volume 27, No. 3, Deitch et al,
30 "Silver Nylon Cloth: In vitro and in vivo Evaluation of
Antimicrobial Activity".

In the last noted reference, the use of a weak direct
current to increase the rate of release of silver ions was
investigated to determine the impact of the increased presence of
35 silver ions. This aspect has been further investigated by several

groups as reported in Antimicrobial Agents and Chemotherapy, February 1976, pp. 357-358, Berger et al, "Electrically Generated Silver Ions: Quantitative Effects on Bacterial and Mammalian Cells"; Antimicrobial Agents and Chemotherapy, November 1976, pp. 856-860, Berger et al, "Antifungal Properties of Electrically Generated Metallic Ions"; American Journal of Veterinary Research, Volume 41, No. 6, pp. 964-966, Colmano et al, "Activation of Antibacterial Silver Coatings on Surgical Implants by Direct Current: Preliminary Studies in Rabbits"; and Plastic and Reconstructive Surgery, Volume 77, No. 3, March 1986, Falcone et al, "Inhibitory Effects of Electrically Activated Silver Material on Cutaneous Wound Bacteria".

The technique of applying a current to a silver coated dressing or purifying devices or medical devices are also disclosed in U.S. Patents 4,291,125; 4,411,648 and published U.K. Patent Application 2,189,677. It is thus apparent that considerable work has been conducted in the field of supplying silver ions in the area of infection or microbial contamination to control and eliminate microbial growth. This concept has been extrapolated further into the field of water treatment, such as disclosed in U.S. Patent 4,407,865 where sand or diatomaceous earth is coated with metallic silver to provide a sterilizing effect as the contaminated waters flow over the filtered material.

It is apparent that sources of silver ions for instance would be particularly useful in surgical and other types of wound dressings. This aspect has been investigated and reported in U.S. Patent 3,800,792, Canadian Patent 1,212,879 and published U.K. Patent Application 2,134,791. Metallic silver is incorporated into the dressing in one form or another and through dissolution silver ions are released into the treated area. U.K. Patent Application 2,134,791 discloses that composites containing various metals, such as silver, gold, palladium, platinum and tin are useful in surgical dressings, where the preferred metal is silver. It is postulated that the slow release of silver ions is facilitated by a galvanic interaction with the moss; i.e., substrate, of the dressing with added metallic or nonmetallic compounds. However, this patent is

- 3 -

silent on how galvanic interaction is developed and directed towards slow release of silver ions in this moss based dressing composition.

European Patent Application 0206024 discloses use of very smooth coatings of various metal combinations on medical devices, such as catheters to provide some antimicrobial activity when the devices are in contact with body fluids.

U.S. Patent 4,418,686 and published U.K. Patent Application 2,194,155A are directed to an implant active in releasing silver ions to treat a bacterial infection. In U.S. Patent 4,418,686, the implant consists of a plurality of spaced-apart metallic bands on a plastic insert where the surfaces of the bands consist of alternate materials, such as silver and gold. The presence of the silver and gold metals in the presence of body fluids results in a galvanic action which is intended to release or liberate silver ions. The implant is of a coiled construction and has metallic bands of considerable size. Such obtained macroscopic galvanic action is not effective, or suitable for most surgical dressings. U.S. Patent 4,252,525 discloses a dental implant where spaced-apart bands of silver and gold are vacuum deposited onto the body of the implant. In addition to these metals, other suitable substances include aluminum, copper, zinc, alloys such as silver-zinc-allantoinate, and silver sulfadiazine, for release of metal ions in providing bactericidal and germicidal action. Other types of implants, which have been treated with silver ions, include catheters which are marketed by Baxter Travenol Laboratories under the trade mark AgX.

The prior techniques involving the use of metal ions in treating microbial infections do not provide a sustained enhanced release of antimicrobial substances. Most existing devices are for short-term applications or suffer from the drawback of very slow release of material. The elements of interest are in fact among the most stable elements. They do not readily dissolve at significant rates on their own, and when in contact with most other metals, will cause such others preferential release. Even when in contact with nobler metals, the differences on the electrochemical scale are quite small that galvanic action occurring over macroscopic areas of

contact does not significantly enhance the release of ions to the level needed.

It is appreciated that the release of metal ions may be expedited by the application of external electric current. However, in many applications, such as in normal bandages or implants, this is practically impossible.

A variety of materials are used every day in treating or preventing infections in humans, animals and the like. For example, catheters, sutures, surgical gloves, implants, bandages, diapers, diaper liners, dressings, small adhesive dressings, sanitary napkins and insoles are just a few. Normally, bandages are used as a barrier to airborne pathogenic organisms infecting a cut or wound. However, once infection occurs, the bandage is no longer of any benefit. If the bandage were provided with a broad spectrum antimicrobial agent, on the portion of the bandage which is in contact with the wound and surrounding skin, the bandage becomes an actively rather than a passively antimicrobial surface or microbial barrier. Catheters, implants, bandages, dressings and other materials, such as above, are used extensively every day by millions of people. As a result, any form of antimicrobial material incorporated into these types of devices must be safe for general unsupervised use, should avoid selection of resistant strains, and should be cost effective. Furthermore, the materials may have to retain their flexibility such as with bandages so as to be readily usable.

It is an object of this disclosure to meet the difficulties encountered in the prior art and to make available both safe and economical actively antimicrobial surface structures and their method of manufacture.

Catheters, implants, bandages, diapers, diaper liners, dressings, and the like can be readily coated with thin films of active elements which, when in contact with body fluids, release substances and ions which stop the growth of or kill various types of microorganisms. As here described, there is no requirement to apply any outside electric current to maintain sustained levels of ion release to treat the infected area.

Disclosure of Invention

In accordance with a first aspect of the invention there is provided on a substrate for use in a biologically dynamic
5 environment, an actively antimicrobial surface film characterized in;
at least a pair of superimposed layers on said substrate,
one of said layers comprising a first element,
the other of said layers comprising a second
electrochemically nobler element than the first element, with the
10 said elements in said layers being in electrical contact,
at least one of the elements being selected from those
which are ionically antimicrobally active and forming ionic solution
in electrolyte containing biological fluid when said fluid is
simultaneously brought into contact with both said layers, the layer
15 of said pair remote from said substrate being substantially
discontinuous thereby permitting access of said biological fluid on
said remote layer to the layer of said pair nearer to said substrate.

In accordance with a second aspect of the invention there is provided a process for the production of an actively antimicrobial
20 surface film on a substrate for use in a biological environment,
which comprises the steps of;

forming a first layer comprising a first element on a
surface of said substrate,

forming a second layer comprising a second element on the
25 first layer, and characterized in;

at least one of said first and second elements being
antimicrobally active and the other element being electrochemically
nobler than said one element,

said elements in said layers being in mutual electrical
30 contact and said one element being ionically soluble in electrolyte
containing biological fluid brought into simultaneous contact with
said layers,

and texturing said layers of said film for permitting said
biological fluid, when brought into contact with the second layer,
35 access through said second layer to the first layer of said film.

In accordance with a third aspect of the invention there is provided on a substrate for use in a biologically dynamic environment, an actively antimicrobial surface characterized in;

a eutectic alloy of elements, one of said elements
5 antimicrobially active and another of said elements being nobler in the electrochemical series than said one element, both said elements being mutually insoluble in solid solution in said alloy, said alloy thereby releasing ions of said one element when said surface is contacted by electrolyte containing biological fluid.

10 In accordance with a fourth aspect of the invention there is provided an actively antimicrobial surface film for a substrate and for use in a biologically dynamic environment characterized in;
a first element dispersed in said film, and
a second electrochemically nobler element in finely divided
15 intimate contact with said first element, at least one of said first and second elements being antimicrobially active, said first element forming ionic solution in electrolyte containing biological fluid when said fluid is brought into simultaneous contact with said two elements.

20 In accordance with a fifth aspect of the invention there is provided a process for the production of an actively antimicrobial surface film on a substrate for use in a biological environment, which comprises the steps of,

forming a layer comprising a first element on a surface of
25 said substrate, characterized in;

said layer being textured,
shadow depositing a second element on the said layer to provide areas of said layer covered by said second element and areas not covered by said second element,

30 at least one of the first and second elements being antimicrobially active and the other element being electrochemically nobler than said one element, said one element being ionically soluble in electrolytic biological fluid brought into contact with said layer.

A variety of ions are active in treating various types of microbial infections or acting as antimicrobials. Some preferred active element ions which have antimicrobial activity are those of platinum, gold, silver, copper, zinc, tin, antimony, and bismuth.

5 The antimicrobial films here described provide, in the presence of an electrolyte, such as provided by body or other biological fluid, an unexpected high reactivity of both the active element and the more noble element, which are released as ions (including ions of the nobler element in some instances, not merely
10 in atomic form) to provide high antimicrobial efficacy.

All the mechanisms of ion release from the films are not yet fully understood, but it is thought that the large surface for galvanic action results, in the presence of electrolyte, in the high reactivity of the substances which are obtained.

15 Both micro-galvanic action and non-galvanic action will contribute to substance release from the film. The element or metal released by galvanic action, namely the one that is less noble on the electrochemical scale, is referred to herein as the "active element". Both the active element and the nobler element are
20 released and contribute to the antimicrobial activity. The enhanced release of the nobler substances or ions from the actively antimicrobial surface film is referred to herein as occurring by "non-galvanic action".

Various embodiments of the invention are exemplified.
25 Although discrete thin layers of the appropriate active elements or metals and more noble elements may be used, it is also evident that alloys, and particularly eutectic alloys, of the various active elements with the nobler elements also work well. The alloy can be of one active element with one nobler element or an alloy of several
30 elements, compounds or metals with at least one or more nobler element.

Eutectic alloys where the active element is insoluble in the solid state and where the other substances or element or elements behave more nobly in the electrochemical sense to the active element
35 can be formed as a single layer actively antimicrobial surface,

because the active element can be brought into solution through galvanic action of biological fluid directly on the eutectic mass.

It is also desirable that biologically active elements such as iodine (for topical activity) be included in the structure of one
5 or more of the layers for release when the matrix breaks up by electrolytic release of the other elements or metals in the film. Such a layer can be of silver/copper/iodine.

By building on the substrate a plurality of layers of a selected active element or metal, in combination with layers of an
10 element metal or alloy more noble in the electrochemical scale than the selective active element, galvanic action is produced in the presence of biological fluids causing release of ions of the active element. For example, silver may be used as the element for building a first layer and then the nobler gold or platinum may be used for
15 the adjacent layer, etc. The preferred nobler elements are selected from the group consisting of platinum, osmium, iridium, palladium, gold, silver and carbon. Since the layers are in contact with one another, electrons readily flow between them, such that, when the layers are in contact with body or biological fluids which carry
20 various salts, galvanic action is set up as miniature or microscopic galvanic cells to release silver ions. Excellent effects can be accomplished for example by using copper in combination with silver, copper in combination with a copper silver alloy, or copper in combination with gold or a silver copper alloy in combination with
25 gold.

The selection of elements determines the ions to be released.

In one preferred embodiment, a thin film structure is realized that releases for example, gold, silver and/or copper for
30 broad spectrum antimicrobial protection. The remaining elements of the preferred active elements group recited earlier are also very effective.

As in the special case for single layer eutectics, with the absence of solid solubility of at least one element of interest
35 discussed above, the thin film morphologies here described for a

nominal film surface area, produce a relatively very large, exposed area of contact between dissimilar elements. This results in an enhanced level of release of elements, due to the creation of plurality of microscopic cells through which enhanced ion release by galvanic action is achieved. The importance of the exposed area of contact of the layer materials in an actively antimicrobial surface film is emphasized since simply having adjacent layers of the appropriate elements with their interfaces buried below the surface, does not produce generally (except in the case of a single eutectic layer) enhanced antimicrobial activity.

Thin or micro-layers of elements can be laid down on a substance by conventional sputtering vapourization, electrochemical multilayer deposition or sol gel techniques. Many of the various thin film deposition techniques, as described in the monograph "Deposition Technologies for Films and Coatings", by R.F. Bunshah, Noves Publications. 1982, can be used to fabricate specifically metallic multilayer films. These include electrochemical and vacuum deposition methods. In the former, the required structures can be realized either by alternate deposition from two separate electrolytic baths, as described by L.M. Goldman, B. Blanpain and F. Spaepen, J. Appln. Phys. 60, 1374 (1986), or by pulsed electroplating from a single bath containing ions of the two metals of interest, as described in U.S. Patent 4,652,348 to J. Yahalom and O. Zadok.

Among the vacuum techniques, the preferred processes are sputtering and evaporation, the latter using resistive, induction or electron-beam heating, since these are most suited to the high rate, large area deposition of metallic films of excellent quality. Multilayers can be realized in a batch process, appropriate for coating discrete substrates individually, wherein the article to be coated rotates or oscillates under two target sources of the elements of interest to be deposited that are shielded from one another so that the article alternately intercepts the vapour stream from one and then the other. Continuous multilayer coating can be realized in a coil to coil operation suitable for flexible substrates, by having the substrate web translate past an array of pairs or series of

sources of the elements. Many such systems are in use, primarily for the deposition onto plastic of solar control films, comprising several layers of different materials, the plastic then being laminated to architectural glass. Metallic multilayer films, with individual layer thicknesses in the range of one to a few hundred nanometers (10 Å to 1000 Å) have been the subject of much recent study due to the variety of interesting mechanical, electronic, and magnetic properties they exhibit for certain metal combinations, including noble metal couples of interest here, as tabulated in Chapter 2 by D.B. McWhan of the monograph "Synthetic Modulated Structures", edited by Chang and Giessen, Academic Press, 1985.

Similarly, ionic sputtering may be used to lay down single elements or a plurality (where two or more definite sources are used). The microlayers, as developed on the substrate, can have a thickness of a few molecules and hence be in the range of 5 to 500 nanometers in thickness. Preferred, however, is a total film thickness of about 1 µ consisting, say, of ten layers each of about 1000 Å thick. The layers are deposited on the surface of the substrate which is to be in contact with the area to be treated; i.e., on a bandage, the layers are applied to the interior surface of the bandage such that when the bandage is applied, the flexible layered film is in contact directly with the body fluids. Since the film is porous the absorbant qualities of the underlying substrate are retained. The substrate need not be flexible however and film may also be deposited on rigid surfaces, such as catheters and implants.

A textured or open film composition can be created in a variety of ways. In a preferred embodiment, where the substrate itself has a rough surface, for example a latex, the layers may be deposited by direct sputtering; that is, substantially at 90° to the surface. Since portions of the rough surface are in effect at an oblique angle to the sputtering beam, irregularities in the deposited surface are created.

When a smooth substrate is used, oblique angle sputtering of the microlayers will result in a suitably textured surface.

Surface structures produced by metal sputtering on rough surfaces and by oblique angle sputtering are described in Current Topics in Materials Science, (1980), Vol. 6, pp. 311-434, by Leamy et al. "The Microstructure of Vapor Deposited Thin Layers" and in
5 Applied Optics, Vol. 23, No. 21, November 1, 1984, pp. 3806-3816 by Guenther "Microstructure of Vapor Deposited Optical Coatings".

These references disclose that surfaces so produced have irregularities which give rise to nodules which grow within the metal laminates. Multiple layers thus become exposed at the surface at
10 these nodules. This multiple exposure of metal interfaces provides for high reactivity of the layered textured film compositions and high antimicrobial efficacy.

Texturing may also be created by etching after deposition of the appropriate multilayers of the film.

15 The texturing of the surface to form a multitude of exposed microlayer interfaces can be generated by a variety of techniques. These include ion beam etching, back sputter etching, chemical etching and mechanical treatment, including, micro abrading, surface cracking by bending, flexing, ultrasonic treatment, expansion of the
20 substrate surface such as by "ballooning" of the article concerned, etc. One preferred method, which produces features on the submicron scale needed, yields uniform texture over large areas, and can be precisely controlled is argon ion beam etching. The development of pronounced texture on materials subjected to ion beam bombardment has
25 been studied extensively, as described in the review article by O. Auciello, J. Vac. Sci. Tech. 19, 841 (1982). Different textures are observed on different materials and these depend also on the parameters of the ion beam used. The mechanism for the induced texture is based on microscopic inhomogeneities, either chemical or
30 microstructural, that are present in the material being etched. As the surface is eroded by sputtering under the action of the ion beam, such regions that may have lower sputtering yield than the surrounding matrix, act as a mask which causes the area around them to recede at a relatively higher rate. Once this localized
35 preferential erosion is initiated, the resulting non-eroded

structures act further to shadow the surrounding regions and so the texture becomes more accentuated as sputtering continues. The argon ions may be supplied from a glow discharge plasma created above the item being etched, which is biased electrically negative so that the positive argon ions are accelerated to it. Alternatively, the argon ions may be generated by broad beam ion sources which have recently become available commercially. The inhomogeneities in the material to be textured can be intrinsic such as impurities and defects. Impurities may also be supplied externally during etching as in the known process of seeded texturing wherein a particular seed material, such as tantalum, is ion deposited at a low rate onto the specimen simultaneously with the sputter etching of the specimen by the ion beam. In the thin film to be etched, an inhomogeneous microstructure can be created in the material during deposition providing both chemical and microstructural modulation. The resulting texture and composition of the etched film is then, in general, dependent on the particular materials in combination and the structure of the layers. Specific textures have been found in such sputtered multilayer films of Ag and Cu as described by M. Tanemura and F. Okuyama in J. Vac. Sci. Tech. A4 (1986). These were studied by them in conjunction with the difficulties that arise from the induced texture in analyzing the multilayer films with ion-based spectroscopic techniques. No consideration, however, was given to potential uses of the texture for its own sake.

Specific embodiments of the invention will now be described with reference to the accompanying drawings.

Brief Description of the Drawings

Figure 1a is a diagrammatic cross sectional representation of a substrate carrying an actively antimicrobial surface of alternate thin layers;

Figure 1b is an enlarged and enhanced section through the layers of Figure 1a showing the peaks and valleys of an ion etched surface;

Figure 2a is a schematic representation in section of a single peak of the etched surface of Figure 1b;

Figure 2b is an enlarged view of the spicules of Figure 1b after shadow cast sputtering of a nobler element on the spicule surface; and

Figure 2c is a further enlarged view of a single spicule of Figure 2b showing the relationship of the elements in the release of the active element ions.

10 Best Modes for Carrying Out the Invention

Figures 1a and 1b illustrate a substrate 20 of material suitable for biological use and which has formed on it several thin layers containing a selected active element alternating with layers containing a nobler element. In this embodiment, a first
15 microlayer 22 comprising either the active element or the nobler element is deposited directly on the contact or outer surface 24 of the substrate. A second microlayer 26 comprising the other element is deposited on the layer 22. Another layer 28 of the same composition as layer 22 is deposited on the layer 26 and a next layer
20 30 is deposited on the layer 28. Though omitted for clarity, further alternated layers continue to be added. Typically, there would be a total of about ten layers each about 1000 Å thick to give a total film thickness of about 1 µ. Each layer is deposited in accordance with standard thin film deposition techniques. To provide a textured
25 surface, the developed layered film of Figure 1a can then be etched, such as for instance by ion etching in accordance with standard techniques to produce in section an arrangement as shown in Figure 1b where several peaks 32 and valleys 34 are formed in the surface and expose layers throughout the film 36. The multiple layers, when
30 exposed on the substrate to body fluids, for example in a wound, provide for release of ions of both elements by galvanic solution of the active element and by non-galvanic solution of the nobler element into the wound area. It is apparent that depending on the make-up of each layer, the corresponding biologically significant elements ions
35 will be released. Each alternate layer need not be of a single

respective element or metal but may be an alloy of two or more elements (each alloy will express a particular electro chemical position with respect to another alloy). Further, each set of layers of one composition may include one or more other elements (such as a halogen, iodine for instance) which will be released as a separate biologically active material when the active element in its layer is released. An example of a preferred structure has layers of silver, copper, iodine, alloy, alternated with layers of silver, the surface layer being preferably of the alloy. If the film is made up of a single eutectic without layering, in which at least one of the elements of interest is insoluble in solid solution, this will release ions without the need for a separate nobler element to be present, the eutectic providing the required elements in a form directly accessible to electrolytic solution.

Other methods of texturing the film surface can be used as heretofore described, such as mechanical working, chemical etching, etc. Each mechanism will produce a characteristic break up of the several layers, however, the principal object in such texturing is to expose the lower layers of the film to biological fluid that will contact the upper surface of the film so that the electrochemical action described can take place more readily.

Figure 2 demonstrates a further development of the textured surface of the antimicrobial system. Figure 2a shows one of the peaks 32 of the active surface 36 of the layered system after etching of Figure 1b. On the surface of each peak 32 is a jagged edge generally designated 38 to indicate a plurality of spicules projecting upwardly of the surface. In Figure 1b, the spicules 40 of the jagged surface 38 are shown in enlarged form. By the technique of shadow deposition, a suitable nobler element as gold or platinum can be directed in the direction of arrow 42 to deposit on one side of the spicules. The spicule constitution will depend upon the layer shown. On the right-hand side of each spicule, a desired nobler metal, say platinum or gold, is deposited. As shown in more detail on Figure 2b and 2c, an individual spicule 40 of peak 32 has a base 43 of, say silver, copper or silver copper alloy, coated on one side

with the more noble element layer 44. Such an individual spicule thus, when exposed to body fluids, sets up an individual microscopic galvanic cell in the region of that spicule to release a continuous supply of the ions of the element or all of the elements in the

5 spicule which are less noble than the element 44. This results in an enhanced metal ion release. Gold may for instance be selected as the more noble element to provide for the galvanic release of the copper or silver ions. However, when gold is desired as the active element ion to be released from a layer, the more noble metal may be

10 platinum. Ions, as well as atoms of the more noble metal, are evidently also released by non-galvanic action. If the microlayers of Figure 1b are alloys of active elements (for instance silver and copper) both Ag and Cu metal ions are released by galvanic action by the arrangement of Figure 2b or 2c when the more noble element is

15 gold. If the alloy is a eutectic even better release can be expected.

The partial coating of a textured surface by oblique sputtering of a different material, to create surface chemical inhomogeneities does not appear to have been exploited elsewhere. The technique of shadow deposition is a conventional one used in the

20 preparation of specimens for transmission electron microscopy to examine the surface topography of a sample. A thin polymeric replica of the surface is produced which is then shadowed by oblique deposition of a thin film, typically of Pt, to create a contrast effect when the replica is viewed in the microscope. The technique

25 is described in Chapter 3 of "Practical Methods in Electron Microscopy", Volume 8, by J.H.M. Willison and A.J. Rower, North Holland Publishing Co., 1980. This demonstrates the ability to coat surfaces controllably having the fine texture of interest here described without overcoating the whole surface and covering over the

30 desired metal interfaces. This may be readily accomplished in production roll coating, for example, by appropriate choice of source position relative to the moving web and the use of slits to define the coated area.

The actively antimicrobial surface film made up of the

35 textured layers of elements provide for enhanced release of ions of

- 16 -

both active element and nobler element, as well as atoms of the nobler element to provide antimicrobial action. If other biologically desirable elements or compounds are included in the active element layer they also will be released. Although the preferred embodiments of the invention have been described with respect to surgical dressings, bandages and the like, the same principles may be applied to other types of surfaces used in microbial treatments, such as water purification and killing of microorganisms in various fluids such as blood. By the technique of texturing, a far more active antimicrobial surface is provided than has been accomplished in other forms of known uses of surface metals in surgical dressings and the like, including those in which macro-galvanic action has been implicated.

The significance of surface texturing of the element microlaminates in producing enhanced release of metal ions, and hence antimicrobial efficacy, is seen from the results in Tables I and IV.

Antimicrobial effects were determined as zones of inhibition measured as in Example I.

EXAMPLE I

Petri plates of agar mixture were prepared according to the manufacturer's (Gibco) instructions, using 1.5% Eagles medium, agar and serum being added to the medium at 50°C. Agar plates were allowed to surface dry for 24 hours. The desired organism, e.g. *S. aureus* ATCC No. 25923 in TSB (Trypticase Soy Broth Difco) was incubated at 37°C for 16 hours then 1 ml transferred to 10 ml TSB for 16 hours; 0.1 ml aliquots of culture dispensed per petri plate and spread to provide bacterial lawn.

The material or disc to be tested was placed on surface of agar, the plates were incubated at 37°C for 24 hours and the zone of inhibition measured and corrected zones of inhibition calculated.

The data shown in Tables I and II were obtained as in Example I. Table III experiment with various organisms was similarly conducted.

The data of Table IV were obtained as in Example II.

35 EXAMPLE II

EXAMPLE II

Sterile synthetic urine was prepared as in Nickel et al, Eur. J. Clin. Microbiol., Vol. 4, #2, pp. 213-218. Discs (coated surfaces or controls) were immersed in synthetic urine at 37°C for various time periods.

Discs were removed from the urine, washed with sterile deionized water and placed on bacterial lawns prepared as in Example I.

Antimicrobial activity was determined as in Example I.

TABLE I

Effect of Various Surface Treatments
on Growth of Staphylococcus aureus

15	<u>Sample</u>	Corrected Zone of Inhibition* <u>(mm)</u>
	Ag foil	<1
20	Ag wire mesh	2.5
	Ag on glass covered 50% with Pt 1 mm bands	<1
	Ag bands and Pt bands	0
	Ag 200 A layers on 1000 A Pt	5
25	on silicone rubber tubing	
	Ag 2000 A layers on 1000 A Pt on silicone rubber tubing	4
	Ag 200 A layers on 1000 A Pt on glass tubing	4

30

*Corrected zone of inhibition = Total zone of inhibition - diameter
of antimicrobial disc

35

TABLE II

Effect of Texturing on Growth of Staphylococcus aureus

	<u>Sample</u>	<u>Texturing</u>	<u>Corrected Zone of Inhibition*</u> <u>(mm)</u>
5	1a	Ag on glass, 3000 A	-
		<1	
	1b	Ag on glass, 3000 A	sputter etch
		<1	
10	2a	Cu on glass, 3000 A	-
		4.5	
	2b	Cu on glass, 3000 A	sputter etch
		4.0	
15	3a	500 A Cu + 500 A Ag layers on glass (10,000 A total)	-
		<1	
	3b	500 A Cu + 500 A Ag layers on glass (10,000 A total)	sputter etch
		10	
20	4a	500 A Cu + 500 A Ag layers on stainless steel (10,000 A total)	-
		3	
25	4b	500 A Cu + 500 A Ag layers on stainless steel (10,000 A total)	sputter etch
		14	
30	5a	500 A Cu + 500 A Ag layers on PTFE (Teflon) (10,000 A total)	-
		3	
	5b	500 A Cu + 500 A Ag layers on PTFE (Teflon) (10,000 A total)	sputter etch
35		13	
	6a	500 A Cu + 500 A Ag layers	

- 19 -

- on latex sheet
(10,000 A total) -
- 11
- 6b 500 A Cu + 500 A Ag layers
- 5 on latex sheet
(10,000 A total) sputter etch
- 16
- 7a 50 A Cu + 50 A Ag layers
on glass (10,000 A total) -
- 10 1
- 7b 50 A Cu + 50 A Ag layers
on glass (10,000 A total) sputter etch
- 14
- *Corrected zone of inhibition = Total zone of inhibition - diameter
15 of antimicrobial disc

TABLE III
Effect of Textured Film on
Composition on Various Organisms

20

Corrected Zone of Inhibition** (mm) on:			
25	<u>Organism</u>	<u>50A Cu and 50A Ag* on latex, not etched</u>	<u>500 A Cu and 500 A Ag* on PTFE, sputter etched</u> <u>500 A Cu and 500 A Ag* on stainless steel sputter etched</u>
	Staphylococcus		
	aureus	13	13 14
	Proteus		
30	mirabilis	13	19 18
	Escherichia		
	coli	1	8 12

* Total thickness was 10,000 A or 1 u

35 ** Corrected zone of inhibition = Total zone of inhibition - diameter
of antimicrobial disc

TABLE IV

Antimicrobial Effect of Textured Films
After Prolonged Exposure to Synthetic Urine

	<u>Days in Synthetic Urine</u>	<u>50 A Cu and 50 A Ag on glass 4 min. etch</u>	<u>50 A Cu and 50 A Ag on latex 4 min. etch</u>
10	0	13	7
	.5	13	
	1	13	7
	2	13	8
15	3	7	8
	4	7	8
	5	5	8

- * Corrected zone of inhibition = Total zone of inhibition - diameter
 of antimicrobial disc
- All experiments were done with *Staphylococcus aureus*
- All were done in synthetic urine, 37°C.

Table I demonstrates the antimicrobial activity of various film surfaces of types comparable to those previously described. A smooth surface of silver alone, as in silver foil, shows no antimicrobial activity. Slight activity is seen when silver wire mesh is tested. The third system of Table I, which would be expected to produce macro-galvanic action at the limited interfaces between the silver and platinum layers, shows no antimicrobial activity. The last three examples of Table I involve smooth metal layers within the range of thicknesses described in European Patent Application No. 0206204, and show only modest antimicrobial activity.

Table II shows the importance of the opening up or texturing of the film microlayers in producing the enhanced antimicrobial efficacy.

- 21 -

Table II shows seven microlayer laminates of silver, copper or alternating layers of silver and copper and compares the antimicrobial activity of these microlaminates with and without texturing. In each case, the untextured microlaminate (a) is not effective as an antimicrobial surface while texturing of the microlaminate, (b) provides a surface having high antimicrobial efficacy.

Table III shows the antimicrobial effect of the textured composition on further bacterial species.

Table IV shows the antimicrobial activity of the textured films after up to five days of exposure to synthetic urine, and shows the persistence of the antimicrobial activity.

Although preferred embodiments of the invention have been described herein in detail, it will be understood by those skilled in the art that variations may be made thereto without departing from the spirit of the invention or the scope of the appended claims.

Industrial Applicability

Medical devices and apparatus such as catheters, implants, bandages, diapers, diaper liners, dressings and the like can thus be provided with surfaces which are actively antimicrobial when in contact with body fluids so as to release ions which inhibit growth of or kill various types of microorganisms.

Claims

1. On a substrate (20) for use in a biologically dynamic environment, an actively antimicrobial surface film characterized in;
at least a pair of superimposed layers (22, 26) on said substrate,
one of said layers comprising a first element,
the other of said layers comprising a second electrochemically nobler element than the first element, with the said elements in said layers being in electrical contact,
at least one of the elements being selected from those which are antimicrobially active and said first element forming ionic solution in electrolyte containing biological fluid when said fluid is simultaneously brought into contact with both said layers, the layer of said pair remote from said substrate being substantially discontinuous thereby permitting access of said biological fluid on said remote layer to the layer of said pair nearer to said substrate.
2. An actively antimicrobial surface film as defined in claim 1, said active element being selected from platinum, gold, silver, copper, zinc, tin, antimony and bismuth.
3. An actively antimicrobial surface film as defined in claim 1, said nobler element being selected from platinum, osmium, iridium, palladium, gold, silver and carbon.
4. An actively antimicrobial surface film as defined in claim 1, at least one of said layers comprising an alloy.
5. An actively antimicrobial surface film as defined in claim 1, comprising a further (28) of said layers on said pair.
6. An actively antimicrobial surface film as defined in claim 1 or claim 5, said one of said layers comprising copper, silver and iodine.

7. An actively antimicrobial surface film as defined in claim 5, each layer being about 1000 A thick.
8. An actively antimicrobial surface film as defined in claim 1 or 5, said layers physically presenting a plurality of spicules (40), each spicule including the layer or layers having said first element and an electrochemically nobler element on said spicule in contact with the last mentioned layer or layers.
9. An actively antimicrobial surface film as defined in claim 8, said nobler element (44) on said spicule being shadow deposited.
10. An actively antimicrobial surface film as defined in claim 1, 5 or 7, said film being textured.
11. An actively antimicrobial surface film as defined in claim 10, an electrochemically nobler element than said first element being deposited on at least part of one of said layers.
12. A process for the production of an actively antimicrobial surface film on a substrate (20) for use in a biological environment, which comprises the steps of;
 - forming a first layer (22) comprising a first element on a surface of said substrate,
 - forming a second layer (26) comprising a second element on the first layer, and characterized in;
 - at least one of said first and second elements being antimicrobally active and the other element being electrochemically nobler than said one element,
 - said elements in said layers being in mutual electrical contact and said one element being ionically soluble in electrolyte containing biological fluid brought into simultaneous contact with said layers,

and texturing said layers of said film for permitting said biological fluid, when brought into contact with the second layer, access through said second layer to the first layer of said film.

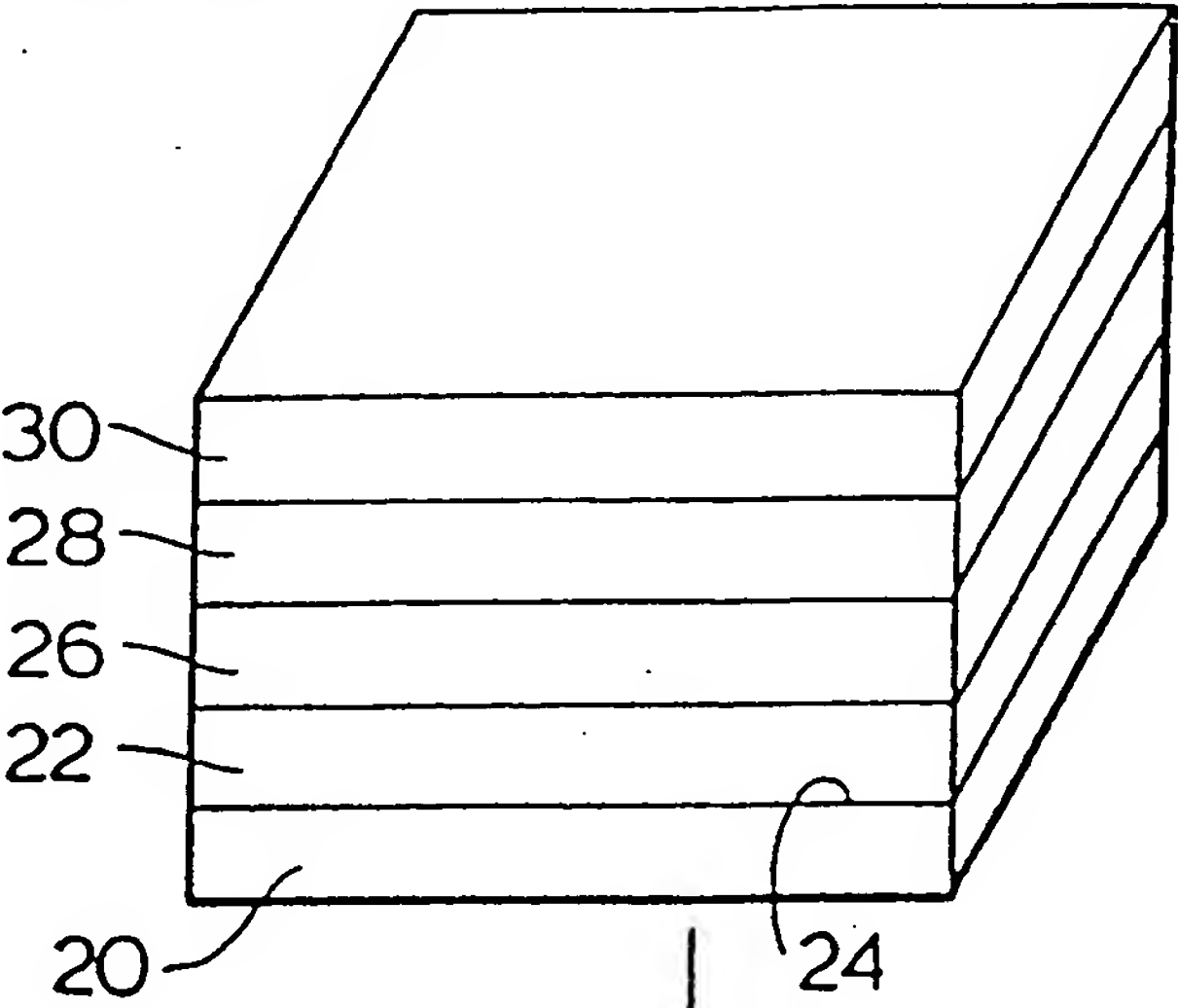
13. A process as defined in claim 12 including the steps of alternately forming additional layers (28, 30) consecutively corresponding in composition to said first and second layers on said second layer to produce a multilayer film.
14. A process as defined in claim 13, wherein the film is about 1 μ and each layer is about 1000 Å thick.
15. A process as defined in claim 12, 13 or 14, said layers being deposited by the step of vapour deposition.
16. A process as defined in claims 12, 13 or 14, said layers being deposited by the step of chemical deposition.
17. A process as defined in claims 12, 13 or 14, said layers being deposited by the step of ionic deposition.
18. A process as defined in claims 12, 13 or 14, including the step of etching said layers for texturing said layers.
19. A process as defined in claims 12, 13 or 14, said one layer being formed by simultaneously depositing a plurality of elements.
20. On a substrate (20) for use in a biologically dynamic environment, an actively antimicrobial surface characterized in;
a eutectic alloy of elements, one of said elements being antimicrobally active and another of said elements being nobler in the electrochemical series than said one element, both said elements being mutually insoluble in solid solution in said alloy, said alloy thereby releasing ions of said one element when said surface is contacted by electrolyte containing biological fluid.

21. An actively antimicrobial surface film as defined in claim 20, said alloy comprising at least three elements.
22. An actively antimicrobial surface film for a substrate and for use in a biologically dynamic environment characterized in;
a first element dispersed in said film, and
a second electrochemically nobler element in finely divided intimate contact with said first element, at least one of said first and second elements being antimicrobially active, said first element forming ionic solution in electrolyte containing biological fluid when said fluid is brought into simultaneous contact with said two elements.
23. An actively antimicrobial surface film as defined in claim 22, said film comprising copper, silver and iodine.
24. An actively antimicrobial surface film as defined in claim 23, said first element being alloyed with another element.
25. An actively antimicrobial surface film as defined in claim 22, said film having a textured surface.
26. An actively antimicrobial surface film as defined in claim 25, said film having been subjected to etching or mechanical treatment for texturing said surface.
27. An actively antimicrobial surface film as defined in claim 22, said first element being selected platinum, gold, silver, copper, zinc, tin, antimony and bismuth.
28. An actively antimicrobial surface film as defined in claim 22, 25 or 27, said nobler element being selected from platinum, osmium, iridium, palladium, gold, silver and carbon.

29. An actively antimicrobial surface film as defined in claim 22, 24, 25, 26, 27 or 28, said nobler element being shadow deposited in said film.
30. A process for the production of an actively antimicrobial surface film on a substrate (20) for use in a biological environment, which comprises the steps of,
forming a layer (22) comprising a first element on a surface of said substrate, characterized in;
said layer being textured,
shadow depositing a second element (44) on the said layer to provide areas of said layer covered by said second element and areas not covered by said second element,
at least one of the first and second elements being antimicrobally active and the other element being electrochemically nobler than said one element, said one element being ionically soluble in electrolytic biological fluid brought into contact with said layer.
31. A process as defined in claim 30, said layer being formed by simultaneously depositing a plurality of elements to form said layer before said shadow depositing step.
32. A process as defined in claim 30 or 31, said layer being formed by vapour deposition.
33. A process as defined in claim 30 or 31, said layer being formed by chemical deposition.
34. A process as defined in claim 30 or 31, said layer being formed by ionic deposition.
35. A medical article comprising the actively antimicrobial surface defined in claim 1, 20 or 22.

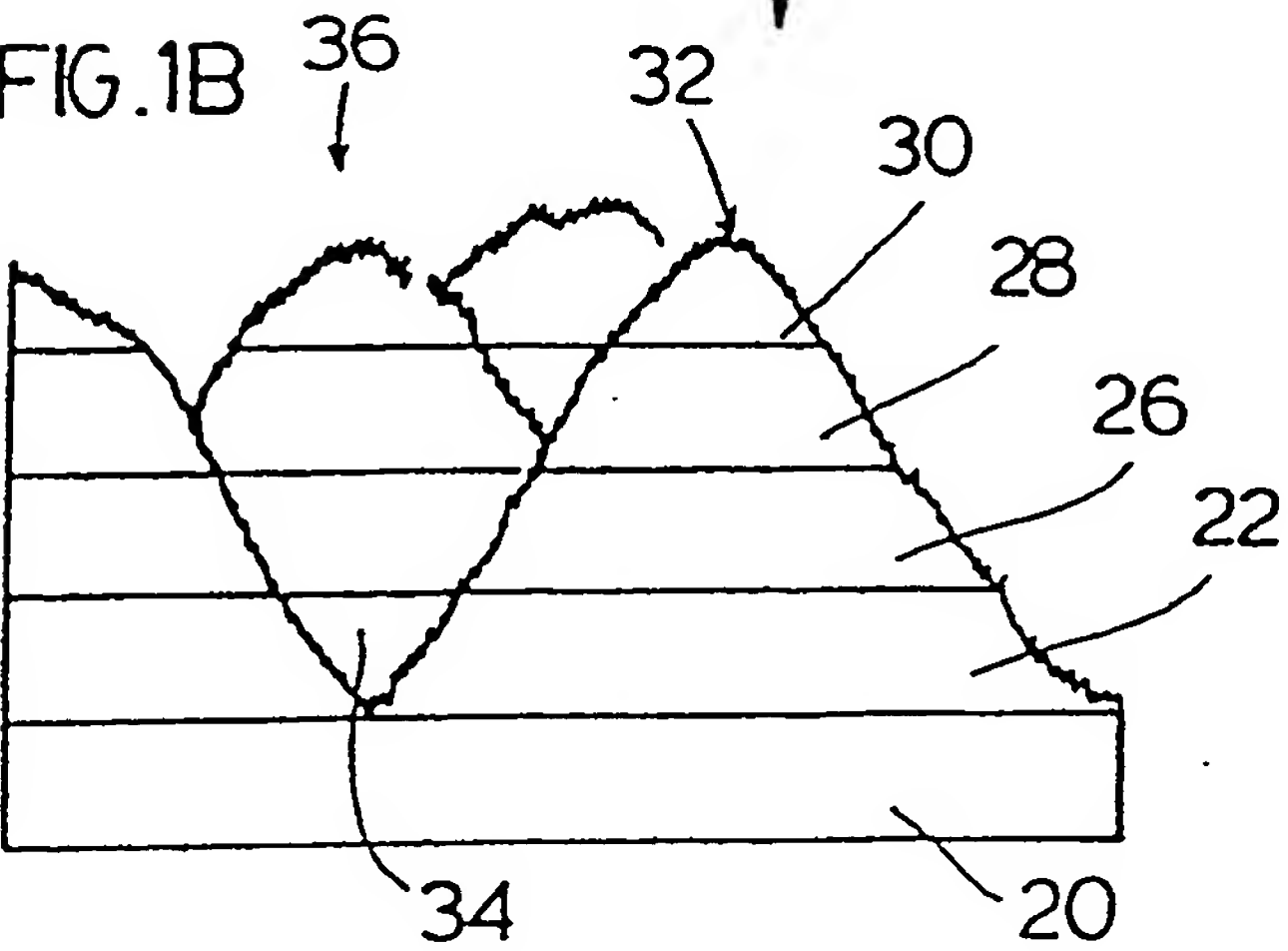
1/2

FIG. 1A

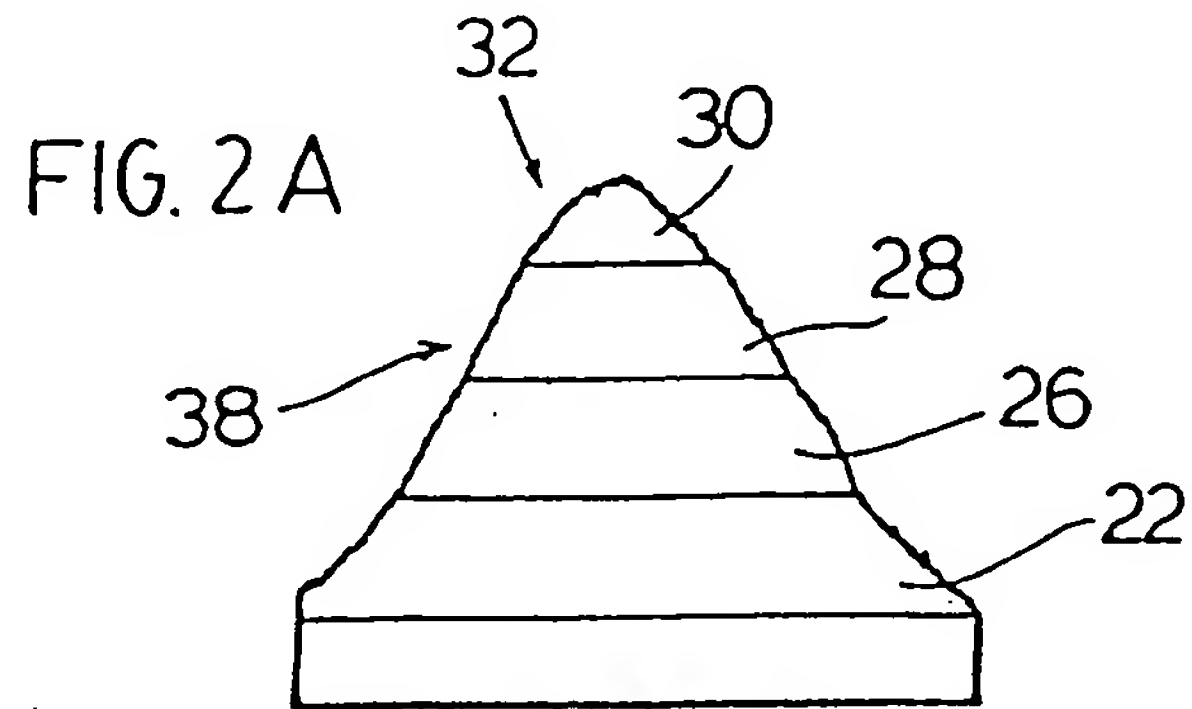


ETCHING

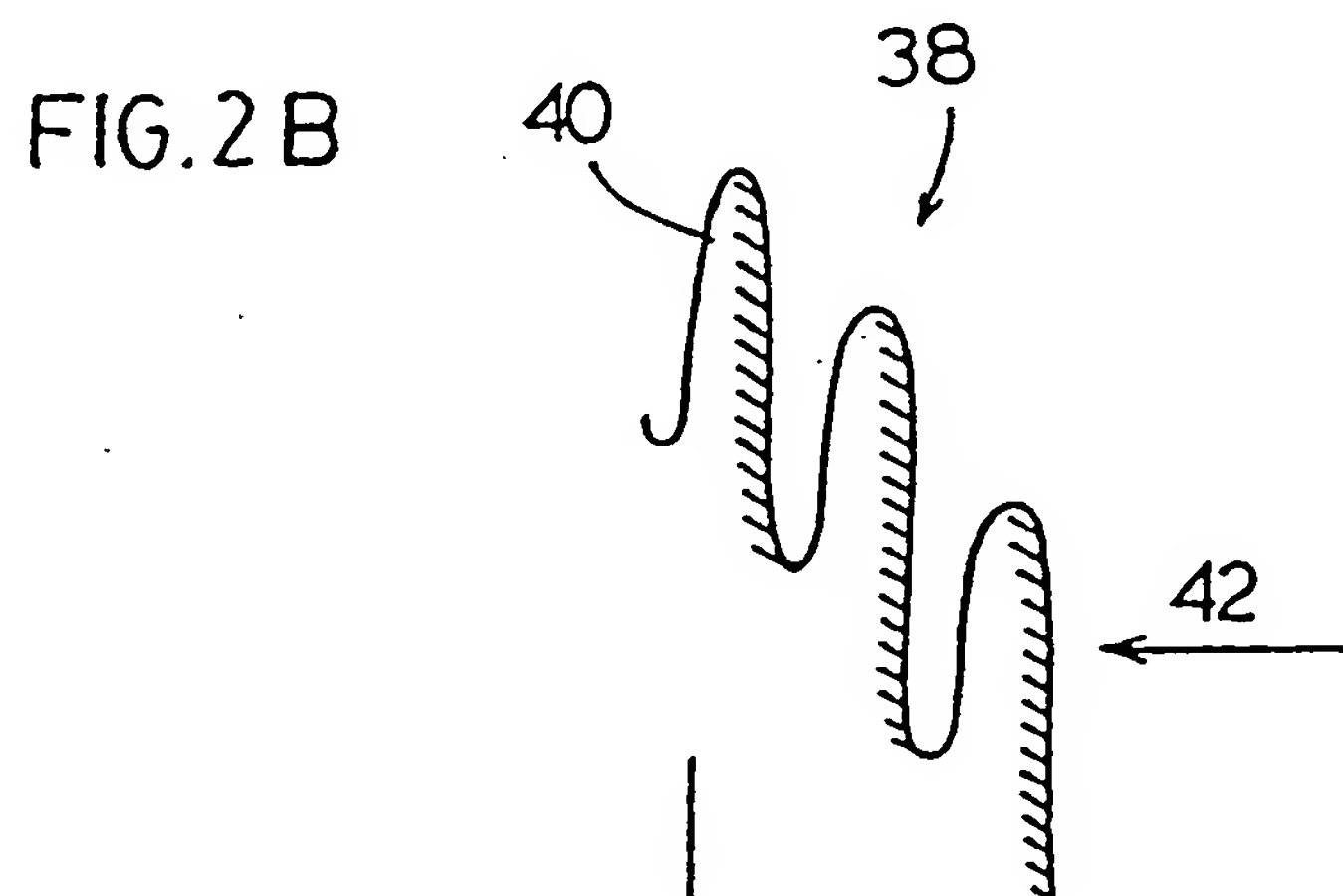
FIG. 1B



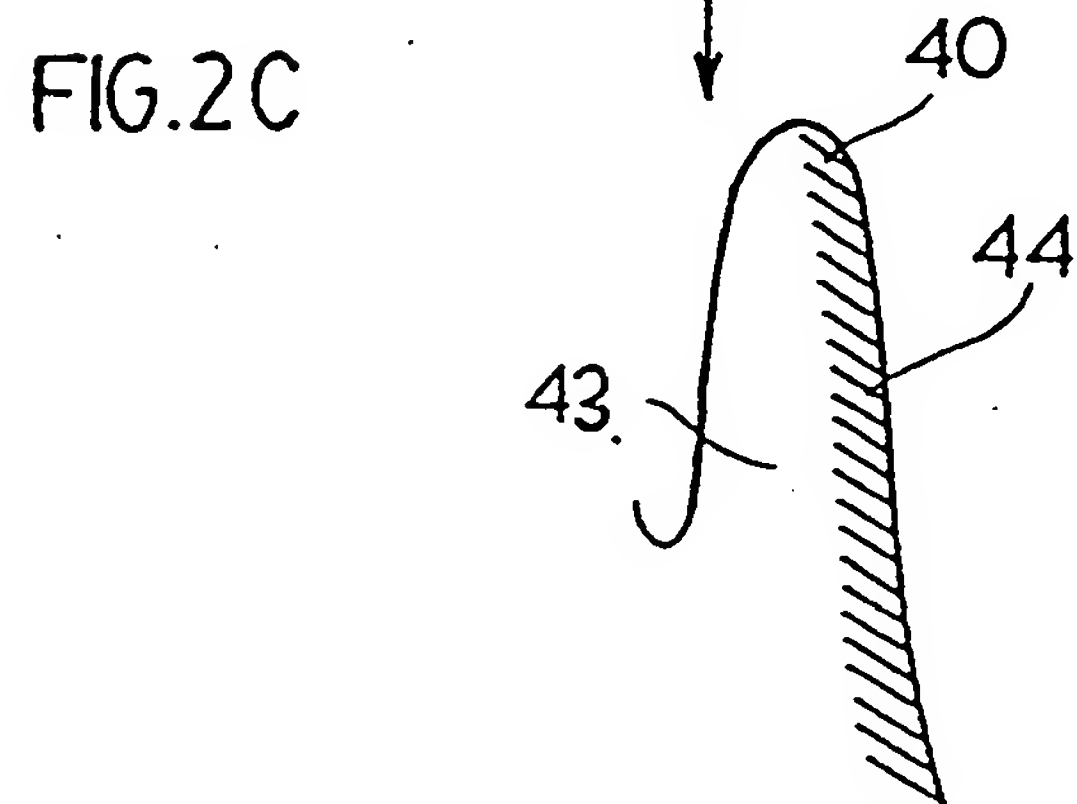
2/2



SHADOW DEPOSITION



ENLARGED



INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 91/00453

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)⁶

According to International Patent Classification (IPC) or to both National Classification and IPC

Int.Cl. 5 A61L29/00; A61L27/00; A61L31/00; A61L15/44
A61M35/00

II. FIELDS SEARCHED

Minimum Documentation Searched⁷

Classification Systems

Classification Symbols

Int.Cl. 5

A61L ; A61M

Documentation Searched other than Minimum Documentation
to the Extent that such Documents are Included in the Fields Searched⁸III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹

Category ¹⁰	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
X	EP,A,0 206 024 (BECTON DICKINSON) 20 July 1988 cited in the application see page 14, line 28 - line 34; claims 1-6,15,16; figures 1-3 ---	1-3,20, 35
X	CH,A,654 738 (I.BLAETTLER) 14 March 1986 see claim 1 ----	1-3,20, 35
A	WO,A,8 102 667 (NATIONAL RESEARCH) 1 October 1981 see page 10, line 14 - line 16 ---	1

¹⁰ Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"A" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search

24 MARCH 1992

Date of Mailing of this International Search Report

10.04.92

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

PELTRE CHR.

ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO.

CA 9100453
SA 54570

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report.
The members are as contained in the European Patent Office EDP file on
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 24/03/92

Patent document cited in search report	Publication date	Patent family member(s)	Publication date	
EP-A-0206024	30-12-86	US-A-	4886505	12-12-89
		AU-A-	3155089	29-06-89
		AU-B-	588598	21-09-89
		AU-A-	5841786	11-12-86
		JP-B-	2005089	31-01-90
		JP-A-	62002947	08-01-87

CH-A-654738	14-03-86	None		

WO-A-8102667	01-10-81	EP-A,B	0048246	31-03-82
		EP-A,B	0048247	31-03-82
		WO-A-	8102668	01-10-81
		GB-A,B	2072514	07-10-81
		GB-A,B	2073024	14-10-81
		US-A-	4476590	16-10-84
US-A-	4615705	07-10-86	-----	